

Primary explosive blast waves cause common types of military-related traumatic injuries. A novel procedure was developed that generates distinct blast waves from 1,3,5-Trinitroperhydro-1,3,5-triazine (RDX), explosive (Zander et al. 2015 *J Neurosci Res* 89:3533). Smith et al. 2016 *Exp Neurol* 286:107). To study direct effects of detonated RDX supersonic blast waves on the brain, we used a novel procedure to expose adult mice in a serum-free medium, covered into a water-filled tank, and RDX assemblies detonated outside the tank. Compared to mock-treated slice cultures, the RDX blasts caused a dose-dependent reduction in pre- and postsynaptic markers. GUR1 was reduced by 25–40% after two consecutive blasts 4 min apart and 25% after a single blast. GUR1 reduction was more pronounced in neurons that were correlated with increased levels of HDAC2, a histone deacetylase implicated in stress-induced reductions in glutamateric transmission and recognition memory (Wei et al. 2016 *J Neurosci* 36:2119). The blast-induced loss of GUR1, NCAM100, and synaptophysin was rapid, with 40–50% reduction in 1 h. The reduction in GUR1 was not observed in neurons that were exposed to a blast and no indication of recovery at 72 h. Interestingly, while protein accumulation events (e.g., tau, TDP-43) are often common features of TBIs (Binnew et al. 2012; Neurosci; Goldstein et al. 2012) *Soc Transl Med*; Smith et al. 2013, *Nature Rev Neurosci*; Zhang et al. 2015) *Cell Death Dis*, the RDX blast waves did not induce any of these markers. The RDX blast waves also induced a reduction in GUR1, which is aligned much closer to synaptic decline profiles from excitotoxic hippocampal slices than to the declines in protein accumulation stress studies. However, the RDX blast waves induced disturbances in synaptic integrity were observed in the absence of calcium-mediated cytoskeletal damage and without evidence of excitotoxicity (Bahr et al. 2002, *Exp Neurol* 174:37). Also, Fluoro-Jade staining found no indication of degenerating neurons in slices exposed to three RDX blasts suggesting that certain levels of military explosives produce a unique type of pathology comprised of altered synaptic integrity before loss of neurons. The RDX blast waves also induced large protein expression explosions, primary blast impacts can cause synaptic compromise without producing overt neurodegeneration, perhaps explaining TBIs cognitive and behavioral changes in blast-induced TBIs sufferers with no detectable neuropathology. The monitoring of individuals' cumulative exposure to military blasts might increase the light of this study. Also, this work will help understand how military blasts might increase the risk of neurodegenerative disease.

Native synaptic density
Green = synaptophysin

Synaptic circuits of the hippocampal network show the different subregions: DG, CA3, and CA1. Additionally, the most widely studied synaptic proteins synaptophysin, GluR1 and synapsin II are illustrated (left). Native synaptic density can be seen in a synaptophysin stained hippocampal slice.

Figure 1 consists of five panels. Panel A is a photograph of a hippocampal slice with several electrodes inserted. Panel B is a histological section showing the locations of the electrodes in the CA1, CA3, and DG regions. Panel C is a photograph of the recording chamber with a plate clamp and pressure sensors. Panel D is a photograph of a pressure sensor recording from a 1.7 g RDX. Panel E is a graph of pressure (pol) vs. time (s) showing a sharp increase at 0.2s.

Hippocampal slice cultures were used to study the effects of realistic blast waves. Slices at culture day 4 are shown distributed on a culture insert membrane (A). The long-term slice cultures display features of the adult brain including intact hippocampal subfields visualized by H & E staining (B; culture day 28). Slice cultures in a six-well plate were sealed in silicon and clamped in position within a warmed, water-filled chamber (C). Three piezoelectric, high-frequency pressure sensors are located directly above the clamped culture plate. A spherical assembly of RDX explosive (D) was detonated outside the chamber and the generated blast wave traveled through the wall of the blast chamber, into the water medium, and was measured as pressure history profile (E). D: dentate gyrus.

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A

SC 1X 2X 3X

GluR1 → 104 83

actin → 49 38

B

100

75

50

25

0

SC 1X 2X 3X

GluR1, percent of control

C

submerge control

RDX blasts

100

75

50

25

0

0 24 48 72

time, h

GluR1, percent of control

D

RDX blasts

100

75

50

25

0

0 24 48 72

time, h

actin, percent

time, h

Military explosive blasts from 1.7-gm RDX assemblies cause a dose-dependent and time-dependent reduction in the AMPA receptor subunit GluR1. Hippocampal slice cultures exposed to 1–3 RDX blasts (1A–1C) were assessed together with mock submerge control slices (SC) by anti-GluR1 immunoblotting (A). Each blot was subsequently stained for actin for a protein load control, and positions for molecular weight standards are shown for 38, 104, and 146 kDa. GluR1 immunoreactivities (means±SE) are shown as percent of levels found in submerge control groups of slices (B) (ANOVA, post-hoc compared to submerge control slices, $^{*}p<0.01$, $^{**}p<0.001$). Other slice cultures were subjected to RDX detonations or mock treatment, then harvested compared to submerge control slices (C) (two-way ANOVA, $^{*}p<0.001$; comparison between control and RDX blast groups at each time point). $^{*}p<0.05$ to assess GluR1 levels across post-blast times (C) (two-way ANOVA, $^{*}p<0.001$; comparison between control and RDX blast groups at each time point). $^{*}p<0.05$ to assess GluR1 levels across post-blast times (C) (two-way ANOVA, $^{*}p<0.001$; comparison between control and RDX blast groups at each time point). $^{*}p<0.05$ to assess GluR1 levels across post-blast times (C) (two-way ANOVA, $^{*}p<0.001$; comparison between control and RDX blast groups at each time point). $^{*}p<0.05$ to assess GluR1 levels across post-blast times (C) (two-way ANOVA, $^{*}p<0.001$; comparison between control and RDX blast groups at each time point).

A

post-blast time:
24h 45h

SC B1-72h

GluR1 →
Synaptophysin →
Synapsin IIIa →
Synapsin IIIb →
actin →

B

Integrated Optical Density

actin GluR1 Synaptophysin Synapsin IIIa Synapsin IIIb

C

SC B1-72h

NCAM 180 →
NCAM 140 →
NCAM 120 →

D (only samples >30% loss in NCAM180)

% change in NCAM isoforms

NCAM 120 NCAM 140 NCAM 180

[illegible]

The graph plots synaptic marker level against days in culture. The Y-axis ranges from 0 to 100, and the X-axis ranges from 0 to 40. A solid black line represents the excitotoxicity condition, starting at 100 and dropping sharply to near zero by day 35. A gray shaded area represents the control slices, starting at 100 and showing a gradual decline. The graph is divided into three regions: 'control slices' (top left), 'excitotoxicity' (bottom left), and 'excitotoxicity' (bottom right).

0 20 25 30 35
days in culture

Figure 3 consists of three panels. The top panel is a Western blot showing GluR1 and HDAC2 protein levels in individual slices. The lanes are labeled 'post-blast time' with '24h' and '48h' markers. The middle panel is a scatter plot showing the correlation between GluR1 and HDAC2 immunoreactivity. The y-axis is 'Immunoreactivity' (0 to 1500) and the x-axis is 'HDAC2' (0 to 2000). A negative linear regression line is shown with the equation $R^2 = 0.373$ and $P = 0.004$. The bottom panel shows two Western blots for pCofilin and pERK2. The lanes are labeled 'BC', 'SC', 'RDX', and 'NIGRA'. The pCofilin blot shows a decrease in pCofilin levels in the RDX lane compared to the other lanes. The pERK2 blot shows a decrease in pERK2 levels in the RDX lane compared to the other lanes.

A synaptophysin DAPI Fluoro-Jade B

control control control

sp sr sr

3X blast 3X blast 3X blast

B 1X blast AMPA

sp sr

C pos SC 3X con AMPA

BDP →

GluR1 →

The results of this study suggest hippocampal slices that receive one or more RDX exposures are functionally compromised. GUR1 and I was further reduced after three consecutive slices.

The presynaptic marker synaptophysin was as ubiquitous as the postsynaptic protein GUR1; 24 h after three detonations and further reduced 48 h post-lash. Interestingly, synaptophysin 24 exhibited a markedly decreased by synaptic deterioration from excitotoxicity versus synaptic decline from the RDX blasts was found to correspond more closely with synaptic deterioration from excitotoxicity versus synaptic decline from accumulation.

abnormal protein accumulation.

RDX detonations disrupt the NMDA-mediated Ca^{2+} pathway that is linked to long-term potentiation and memory encoding.

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Fluoro-Jade staining found no indication of degenerating neurons in slice cultures exposed to three RDX blasts, suggesting that live-exposed explants can produce synaptic alterations in the absence of cellular degeneration.

Together, these results indicate that detonated RDX explosives cause distinct losses of synaptic proteins before cell death, perhaps exposing the underlying mechanisms of blast-induced TBIs with no detectable neuropathology.

Acknowledgements & References

[illegible]

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